

## Ozone against mycotoxins and pesticide residues in food: Current applications and perspectives

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### Abstract

Food safety may be compromised by the presence of chemical contaminants, such as mycotoxins and pesticide residues. Mycotoxins are natural contaminants produced by certain species of filamentous fungi and can cause toxic effects on human health. Pesticide residues are any specified substance in food resulting from the use of a pesticide with toxicological significance. To protect consumers from these toxic substances, different food regulatory agencies have set maximum levels permitted in different raw materials and processed foods. However, recent research has demonstrated a high incidence of both mycotoxins and pesticide residues (not simultaneously) in foods marketed all around the world, sometimes with levels above the regulated limits. One way to reduce such contaminants is to use ozone (O<sub>3</sub>) in food processing. Due to its high potential as an oxidant, O<sub>3</sub> or the radicals generated in the ozonation process react with mycotoxins and pesticide residues that lose their toxicity due to molecular degradation. In this review paper the recent research into using O<sub>3</sub> for gaseous ozonation and ozonized water to decontaminate food by eliminating and/or reducing mycotoxins and pesticide residues are discussed. Also the changes promoted in food quality attributes, the possible formation of degradation products of toxic relevance, as well as some perspectives for the future use of this technology in food processing are explored.

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### Keywords

Food safety  
Chemical contaminants O<sub>3</sub>  
Gaseous ozonation  
Ozonized water

### Introduction

Every day more and more consumers, worldwide, are becoming aware of food safety and the risks associated with its contamination by microorganisms and by toxic compounds (Kher *et al.*, 2013). The presence of contaminants in food, such as pesticide residues or mycotoxins, raises concerns in terms of public health and food safety.

Pesticides are used in agriculture to improve productivity by protecting crops from disease and infestation. However, they must be applied in accordance with the Good Agricultural Practices (GAPs) and the levels present in foods must be below the Maximum Residue Levels (MRLs). MRLs of pesticide vary greatly worldwide, because countries have different requirements and different legal limits (EFSA, 2015; Handford *et al.*, 2015).

Nowadays, the high global usage of pesticides, approximately two million tons per year (De *et al.*,

2014), allied with the increased resistance of pests and pathogens, has posed a renewed concern on the use of pesticide in food (Liu *et al.*, 2015). Despite the surveillance carried out by the competent authorities, recent research has shown significant incidence of pesticide residues at levels above the MRLs in various foods, such as: vegetables (Akoto *et al.*, 2015; Chourasiya *et al.*, 2015), coffee (Oliveira *et al.*, 2016), honey (Bargańska *et al.*, 2013; López *et al.*, 2014) apples (Lozowicka, 2015), guava, kaki and peach (Jardim *et al.*, 2014) and cereal grains (Min *et al.*, 2012; Lozowicka *et al.*, 2014).

The ingestion of food contaminated with pesticide residues is associated with endocrine, reproductive and nervous disorders, as well as the risk of cancer (US EPA, 2014; Blaznik *et al.*, 2015; Chiu *et al.*, 2015). This contamination is of particular concern for infants and adolescents due to their lower detoxification capacity and high intake of food per kg body weight (Lombard, 2014).

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Mycotoxins are other food chemical contaminant with a rising concern in public health. Mycotoxins are harmful metabolites produced by a number of filamentous fungi, such as, *Aspergillus* spp., *Fusarium* spp. and *Penicillium* spp. that may contaminate food in the field and/or when improperly stored (EFSA, 2013; Cheli *et al.*, 2014). Their effects on human health depend on the mycotoxin and the level consumed. These effects may vary from mild intoxication to cancer and death (IARC, 2002; Marroquín-Cardona *et al.*, 2014). GAPs include procedures to prevent infestation by fungi in stored products and avoid consequent contamination by mycotoxins; however, as they are natural contaminants, all humans and animals are exposed to such hazard. Recent studies, using urinary and breast milk biomarkers, carried out all over the world, showed a high human exposure to different mycotoxins, such as aflatoxins, ochratoxin A, zearalenone and deoxynivalenol (Srey *et al.*, 2014; Iha *et al.*, 2014; Rubert *et al.*, 2014; Gerding *et al.*, 2015).

Recent research has demonstrated that ozonation can be used to decontaminate and remove mycotoxins and pesticide residues in food, especially in fresh fruits, vegetables and grains. The molecular ozone ( $O_3$ ) or the hydroxyl radicals generated in the process, especially in ozonized water, react with these contaminants promoting their degradation and form lower molecular weight products, thus eliminating or reducing the biological activity of these contaminants in terms of toxicity (Ikehata *et al.*, 2006; Diao *et al.*, 2012; Luo *et al.*, 2014a). The United States Food and Drug Administration (FDA) has recognized, since 2001, that  $O_3$  is GRAS (Generally Recognized as Safe) for the treatment, storage and processing of food and water (FDA, 2001). The advantages of using  $O_3$  to decontaminate food products over other oxidants is that it is environmental friendly to produce and its use does not leave any residues in the food, as the  $O_3$  dissociates into oxygen. Consequently it is recognized as a “green technology” (O’Donnel *et al.*, 2012; Greene *et al.*, 2012).

Ozonation efficacy to degrade mycotoxins and pesticide residues depends on the  $O_3$  concentration, exposure time, type of food, moisture content, mode of application (gas or water), among others factors. Moreover, ozonation may cause positive changes in the quality of the food, such as, increasing the volume of breads and cakes or increasing strength and clarity of flours (Caballero *et al.*, 2007; Li *et al.*, 2012). However, as it is not a universally beneficial process,  $O_3$  can also cause negative alterations, such as oxidation of lipids, changes in sensory characteristics, color loss, degradation of phenolic

compounds and some vitamins, among other adverse effects (Patil *et al.*, 2010; Gabler *et al.*, 2010). These negative effects must be studied in greater depth to define the limitations of this technology.

This paper reviews the most recent studies on the reduction of mycotoxins and pesticide residue levels promoted by ozonation, including changes in the quality of the food due to its treatment. The possible formation of degradation products of toxic relevance and the future prospects for scientific research and the industrial use of  $O_3$  in the food science and technology field are also discussed.

#### *Industrial production of $O_3$ and legislation for food processing*

Ozone must be produced on-site for immediate use in ozonation processes due to its instability and rapid dissociation into  $O_2$ . When  $O_3$  is used to decontaminate food, it is usually produced with ozonizers based on corona discharge. Ozonizers expose  $O_2$  molecules to a high voltage electrical discharge which initiates the formation of free radical oxygen and thereby generates  $O_3$  (WCBBC, 2006). The corona discharge method can obtain high concentrations of  $O_3$  at a low cost; however, UV radiation can also be used for commercial production of  $O_3$  but with a lower concentration and yield (Tapp and Rice, 2012).

Ozone can be applied in the gaseous form directly into the food, as occurs in cereal grains, or it can be bubbled into water to produce ozonized water, which is especially suitable for the raw materials that require an aqueous disinfection step (Coelho *et al.*, 2015). In the gaseous form, the half-life of ozone is a few hours in the presence of food, however, in still air at 0% humidity it can have a half-life of up to 25 h (McClurkin *et al.*, 2013). When bubbled in water,  $O_3$  dissolves partially, forming hydroxyl radicals (OH $\cdot$ ) that can oxidize the contaminants more efficiently than molecular  $O_3$  (Takahashi *et al.*, 2007). Ozonized water can be used for washing a variety of foods, such as fruits and vegetables, and it can even be used on cereal grains that require water as a conditioning step (tempering) prior to the milling process, like wheat grains (Ibanoglu, 2001).

There are no concentration limits for the application of  $O_3$  in food; however, as it is a GRAS substance its concentration should be as low as reasonably achievable (ALARA) and also in accordance with the Good manufacturing practices of the food industry. The American conference of governmental industrial hygienists set a limit of 0.2 mg/m $^3$  of  $O_3$  exposure for an 8-hour-day (FDA, 2014), while the World Health Organization (WHO)

recommends 0.1 mg/m<sup>3</sup> for an 8-hour mean (WHO, 2006). As O<sub>3</sub> is a toxic gas, levels higher than these limits may cause undesirable physiological effects on the central nervous system, heart, and vision (PubChem, 2015).

#### Current concerns related to mycotoxins and the use of O<sub>3</sub> to reduce food contamination

The contamination of food with mycotoxins is a serious public health concern and it can lead to many health problems due to the diverse toxic effects promoted by these substances, such as, cytotoxicity, genotoxicity, immunotoxicity, carcinogenic or teratogenic effects (Stoev, 2015). The main mycotoxins related to food contamination are the aflatoxins, fumonisins, zearalenone, citrinin, patulin, ochratoxin A, deoxynivalenol and other trichothecenes (Rocha *et al.*, 2014; Wu *et al.*, 2014).

In cereal grains, *Fusarium* toxins, such as trichothecenes, zearalenone and fumonisins, are the most commonly detected, particularly in the pre-harvest phase (De Ruyck *et al.*, 2015). Trichothecenes, like deoxynivalenol (DON), are known for their strong capacity to interfere with protein synthesis and induce immunosuppression (Antonissen *et al.*, 2014). Fumonisins are associated to esophageal cancer and can also interfere in the biosynthesis of sphingolipids with consequent cell activity disorders. Zearalenone is a potent estrogenic metabolite and can cause infertility, abortion and other reproduction problems (Yazar and Omurtag, 2008).

*Aspergillus* spp. and *Penicillium* spp. fungi are of great importance during food storage. If adequate moisture and temperature conditions exist, *Aspergillus* spp. can produce mycotoxins, such as aflatoxin, especially in oilseeds and cereals (Gorayeb *et al.*, 2009). Aflatoxins (AF) are one of the most important environmental toxins and the AFB<sub>1</sub> is the mycotoxin with the most toxicity in this group, with hepatocarcinogenic and immunosuppressive activities (Magnussen, 2013). Both *Aspergillus* spp. and *Penicillium* spp. may produce ochratoxins that mainly contaminate cereals, but they can also contaminate grapes and their derivatives, like wine. Ochratoxin A is the most relevant in this group and has been reported to be nephrotoxic and carcinogenic to humans (Sorrenti *et al.*, 2013). These fungi may also produce patulin in apples, which is a genotoxic and cytotoxic substance (Glaser and Stopper, 2012). Certain species of *Penicillium*, *Aspergillus* and mainly *Monascus* may produce citrinin in rice, which has nephrotoxic, hepatotoxic and carcinogenic activity (Li *et al.*, 2012b). Figure 1 illustrates the molecular structure of the main mycotoxins that occur in food.

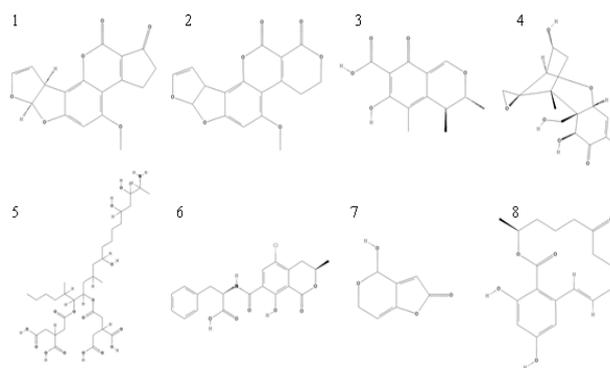


Figure 1. Molecular structures of the main mycotoxins found in food. 1- aflatoxin B1, 2- aflatoxin G1, 3- citrinin, 4- deoxynivalenol, 5- fumonisins B1, 6- ochratoxin A, 7- patulin, 8- zearalenone. Reference: Adapted from PubChem (2015).

The production of mycotoxins depends on the environmental conditions during plant growth and subsequent food storage, consequently their presence is sometimes unavoidable in food (Stoev, 2015). Mycotoxins are stable to traditional industrial processes applied to raw materials; thus if the raw food is contaminated, these mycotoxins will also be present in the processed foods, posing a health risk to consumers (EFSA, 2013; Tibola *et al.*, 2015).

When O<sub>3</sub> is applied at low concentrations to the storage of fruits and cereals with a long exposure time it can control or inhibit growth, germination and sporulation of fungi, thus preventing the production of their toxins (Giordano *et al.*, 2012; Feliziani *et al.*, 2014; Hansen *et al.*, 2013). However, these effects are very dependent on the fungal species, growth stage, O<sub>3</sub> concentration and exposure time (Freitas-Silva and Venâncio, 2010).

On the other hand, to promote the molecular degradation of mycotoxins, high concentrations levels of O<sub>3</sub> are needed. The exposure time, type of food, moisture content and temperature are also factors that directly affect the efficacy of this decontamination (Ikeura *et al.*, 2011; Li *et al.*, 2014). The degradation rates of mycotoxins by O<sub>3</sub> reported in the literature vary widely due to the different experimental conditions used in each study. Table 1 summarizes some recent experimental studies involving mycotoxin decontamination by ozonation in different kinds of foods, including the conditions applied and the degradation percentage obtained.

Besides experimental laboratory studies, McDonough *et al.* (2011) evaluated the use of ozonation on a commercial scale by applying O<sub>3</sub> at 4.7% in air to corn kernel using a continuous-flow system. The O<sub>3</sub> was delivered into a screw conveyor with retention time for the grains moving through the system equal to 1.8 min. Under these conditions

Table 1. Summary of some recent studies involving mycotoxin decontamination in food by O<sub>3</sub>

Food	Mycotoxin	Conditions	Reduction levels (%)	Reference
Peanut	AFT AFB <sub>1</sub>	6.0 mg of O <sub>3</sub> /L for 30 min	65.8% 65.9%	Chen <i>et al.</i> (2014)
	AFB <sub>1</sub>	50 mg of O <sub>3</sub> /L for 60 h	89.4%	Diao <i>et al.</i> (2013)
Brazil nut (in-shell)	AFT AFB <sub>1</sub>	21 mg of O <sub>3</sub> /L for 96 h	30% 25%	Alencar <i>et al.</i> (2012)
	AFT	14 mg of O <sub>3</sub> /L for 30 days	100%	Giordano <i>et al.</i> (2010)
Corn flour	AFB <sub>1</sub> AFG <sub>1</sub> AFB <sub>2</sub>	75 mg of O <sub>3</sub> /L for 60 min	78.7% 73.7% 70.6%	Luo <i>et al.</i> (2014b)
	Dried figs	13.8 mg of O <sub>3</sub> /L for 180 min	95.2%	
Flaked red pepper	AFB <sub>1</sub>	Ozonized water (1.71 mg of O <sub>3</sub> /L) for 180 min	88.6%	Zorlugenç <i>et al.</i> (2008)
	AFB <sub>1</sub>	66 mg of O <sub>3</sub> /L for 60 min	93%	Inan <i>et al.</i> (2007)
Pistachio	AFB <sub>1</sub> AFT	9 mg of O <sub>3</sub> /L for 420 min	23% 24%	Akbas and Ozdemir (2006a)
	AFB <sub>1</sub>	0,004% of O <sub>3</sub> in air for 20 min	96.6%	El-Desouky <i>et al.</i> (2012)
Wheat grains	AFB <sub>1</sub> AFB <sub>2</sub> Citrinin	O <sub>3</sub> at 60 µmol/mol/ for 180 min	96.6% 84.5% 75.27%	Savi <i>et al.</i> (2014b)
	DON	O <sub>3</sub> at 60 µmol/mol for 180 min	100% <sup>1</sup>	Savi <i>et al.</i> (2014a)
Apple juice	Patulin	12% of O <sub>3</sub> (w/w) bubbled into juice	100% <sup>1</sup>	Cataldo (2008)

<sup>1</sup> Lower than the Limit of Detection (LOD) for the method used. AF: Aflatoxins. AFT: Total aflatoxins. O<sub>3</sub>: Ozone.

there was an approximate reduction of 30% in the aflatoxins.

Some studies have demonstrated the efficacy of O<sub>3</sub> to degrade mycotoxin standards in solution, which is also a method that helps explain how O<sub>3</sub> promotes the mycotoxin degradation. Young *et al.* (2006) studied the effects of ozonized water at 25 ppm in the degradation of DON, nivalenol (NIV) and other trichothecenes. These authors concluded that the degradation begins with attack of the C9-10 double bond by O<sub>3</sub> causing the mycotoxin to breakdown into organic acids, aldehydes, and ketones. This process effectively reduced the trichothecenes levels in solution. Dudziak (2012) studied zearalenone degradation by ozonized water at 1 mg/L for 20 min. This treatment reduced the concentration of the mycotoxin to undetectable levels. The authors concluded that the use of a high exposure time contributes to a more effective degradation. Freitas-Silva (2011) also demonstrated the potential of ozonized water at 20 mg/L to degrade a solution of cyclopiazonic acid. The author point out that the ozonized water can decontaminate not only mycotoxin standards with efficacy but also raw materials, laboratory equipment and reagents for disposal.

In order to study the practical use of O<sub>3</sub> treatment, besides the proven reduction of mycotoxins levels that

can be verified by analytical techniques, it is essential to identify the molecular products formed by the degradation of mycotoxins, and to have knowledge of their toxicity. Luo *et al.* (2014a) identified six degradation products of aflatoxin B<sub>1</sub> by ozonation in aqueous solution at different O<sub>3</sub> concentrations, through the structure–activity relationship, authors confirm that the ozonation eliminated the toxicity of aflatoxin B<sub>1</sub>. According to these authors, the unsaturated molecules are more easily attacked by O<sub>3</sub>, while the saturated ones are more resistant to detoxification. Diao *et al.* (2012) reported that the oxidation of AFB<sub>1</sub> in acetonitrile, using gaseous ozone at 6.28 of O<sub>3</sub> mg/L, formed thirteen degradation products and it eliminated the molecular structure of AFB<sub>1</sub> responsible for its toxic effects. According to the authors, the toxicity of aflatoxin was significantly reduced because of the disappearance of the double bond on the terminal furan ring or the lactone moiety on the benzene ring.

Another way to verify if ozonation can eliminate or significantly reduce the toxic effects of food contaminated by mycotoxins was demonstrated using animal models. Diao *et al.* (2013) showed that peanuts contaminated with AFB<sub>1</sub> and then ozonized at 50 mg of O<sub>3</sub>/L for 60 h did not cause any symptoms of toxicity or changes in the appearance and behavior of female Wistar rats. These authors showed that

the deleterious effects of AFB<sub>1</sub> were reduced by the ozonation to such an extent that the health of the animals was not affected. Gaou *et al.* (2005) investigated if the treatment of wheat grains with 5 g of O<sub>3</sub>/kg grains would promote adverse effects on the health of Dark Agouti rats due to grain consumption. After 4 weeks, clinical, hematological, biochemical blood, urinary and histopathological determinations were not significantly affected; thus the consumption of the ozonized grains was considered safe.

Luo *et al.* (2014c) studied the toxicity of the degradation products from AFB<sub>1</sub> formed in artificially contaminated ozonized corn using the human hepatocellular carcinoma cell line (HepG2) as the model. The authors reported that the toxicity of ozone-treated corn had no significant difference with the corn free of mycotoxins. These different techniques indicate that the gas ozonation process and the use of ozonized water are effective to reduce food contaminated by mycotoxins and that they do not produce degradation products of any known relevant toxicity.

#### *Applications of O<sub>3</sub> to reduce pesticide residues in food*

Pesticide residues are any specified substance in food, agricultural commodities, or animal feed resulting from the use of a pesticide, including its conversion products, metabolites, reaction products and impurities considered to be of toxicological significance (FAO, 2001). The impact on human health due to the use of pesticides in agriculture is of increasing concern in the eyes of the public due to the evidence between human exposure to these residues and chronic diseases, such as cancers, diabetes, neurodegenerative, as well as birth and reproductive disorders (Mostafalou and Abdollahi, 2013; Parrón *et al.*, 2014).

Food containing pesticide residues is a direct source of exposure and although industrial or domestic processing of food, such as peeling and cooking, can help reduce the contamination, these residues will partially remain in the final product, representing a health risk if ingested in high concentrations (Kaushik *et al.*, 2009). The MRLs for pesticides in food samples are regulated throughout the world, and are basically concerned with the quality, efficacy and safety in the use of pesticides; however, there is not a global harmonized legislation (Malik *et al.*, 2010). In general, developed nations have more stringent regulations than developing countries, which lack the resources and expertise to adequately implement and enforce regulation, posing a technical barrier to

trade and to public health protection (Handford *et al.*, 2015).

In recent years, powerful analytical methods, especially mass spectrometry, have played a vital role in the identification and quantification of these substances in a variety of matrices (Malik *et al.*, 2010; Romero-González, 2015). These up-to-date tools have identified pesticide residues in human blood, urine, breast milk and hair by various authors, indicating a high level of exposure to humans worldwide (A El-Morsi and Rahman, 2012; Dewan *et al.*, 2013; Yusa *et al.*, 2015).

The number of studies investigating the potential use of ozonation to reduce pesticide residue levels in food has increased in recent years, especially studies concerning ozonized water to wash vegetables and gas ozonation to treat cereals. The degradation of these pesticide residues can be carried out via the molecular O<sub>3</sub> reaction pathway with the food, which gives rise to selective reactions, especially with unsaturated and aromatic hydrocarbons; or, by an indirect pathway, involving radicals of higher oxidation potential that can attack organic and inorganic molecules with non-selective reactions (Ikehata *et al.*, 2006; Ormad *et al.*, 2008).

For the treatment of drinking water, more drastic conditions of ozonation in terms of concentrations and exposure time may be applied compared with the treatment for food. Lafi and Al-Qodah (2006) studied ozonation associated with UV radiation for the treatment of drinking water and reported a 100% degradation of the insecticide deltamethrin and also an 80% reduction of halogenated and non-halogenated compounds. Ormad *et al.* (2008) investigated the reduction of 44 pesticides by chlorine or O<sub>3</sub>. These authors concluded that ozonation was more efficient than chlorine as it eliminated about 70% of the pesticides studied, and with an advantage over chlorine as it did not form trihalomethanes after the treatment. When these authors incorporated a carbon activated absorption step together with ozonation, the levels of pesticides in water were reduced by 90%, demonstrating the high potential of O<sub>3</sub> when applied together with other technologies in the treatment of drinking water.

Ikeura *et al.* (2011; 2013) studied the application of ozone microbubbles (<50 µm in diameter) in water at low O<sub>3</sub> concentrations (1 to 2 ppm) to wash fruits and vegetables, concluding that the use of this technique is highly effective in quickly removing pesticide residues. According to the authors, the microbubbles allowed the O<sub>3</sub>, which is highly insoluble in water, to be dissolved easily, generating higher amounts of hydroxyl radicals, which are very

Table 2 - Summary of recent studies involving pesticide residue degradation in food by O<sub>3</sub>

Food	Pesticide residues	Experimental conditions	Reductions obtained	Reference
Wheat grains	Fenitrothion	O <sub>3</sub> at 60 µmol/mol for 180 min	66.7%	Savi <i>et al.</i> (2015)
	Deltamethrin		89.8%	
White cabbages	Chlorfluazuron	Ozonized water exposed to 250 mg of O <sub>3</sub> /h for 15 min	60%	Chen <i>et al.</i> (2013)
	Chlorothalonil		55%	
Persimmon leaves	Fenitrothion	Microbubbles in water at 2 ppm of O <sub>3</sub> for 15 min	56%	Ikeura <i>et al.</i> (2013)
	Benomyl		50%	
Citrus fruits	Chlorothalonil	10 ppm of O <sub>3</sub> for 5 min	100%	Kusvuran <i>et al.</i> (2012)
	Tetradifon		98.6%	
	Chloropyrifos		94.2%	
Lettuce	Fenitrothion	Microbubbles in water at 1 ppm of O <sub>3</sub> for 5 min	58%	Ikeura <i>et al.</i> (2011)
Table grapes	Fenhexamid	Fumigation for 1 h (10 mL of O <sub>3</sub> /L)	68.5%	Gabler <i>et al.</i> (2010)
	Cyprodinil		75.4%	
	Pyrimethanil		83.7%	
	Pyraclostrobin		100.0%	
<i>Brassica rapa</i>	Diazinon Parathion Methyl-parathion Cypermethrin	Ozonized water using 1.4 to 2.0 mg of O <sub>3</sub> /L for 30 min	Reduction from 60 to 90%	Wu <i>et al.</i> (2007)

effective at decomposing organic molecules.

In contrast to its use for drinking water, ozonation of food is limited by the undesired changes that occur in the quality of these due to the high oxidizing capacity of O<sub>3</sub>. Consequently, an optimization of the conditions for decontamination must be studied for each food and for each pesticide residue. Table 2 summarizes some recent studies involving pesticide residue degradation in food through ozonation.

#### Food alterations due to ozonation treatment

When O<sub>3</sub> is used during storage or food processing in order to reduce the levels of residues or contaminants, its high oxidation power may promote unwanted changes in the food quality. Fruits and vegetables are the most affected by the negative effects of ozonation due their high moisture content, enzymes and phenolic compounds. Patil *et al.* (2010) bubbled apple juice with 0.048 mg of O<sub>3</sub> for 10 min and observed a change of color and reduction of phenolic compounds. These authors suggested that the O<sub>3</sub> and the hydroxyl radicals (OH<sup>-</sup>) generated may have opened the aromatic rings of the phenolic compounds which lead to the oxidation of organic acids, aldehydes and ketones. They also suggested that the loss of color was a consequence of the breakdown of conjugated double bonds. Gabler *et al.* (2010) also reported color changes and other injuries when they used ozonized water at 5 mg of O<sub>3</sub>/L to wash grapes for 1 h. However, such changes

can also be promoted by other oxidant processes, for instance; loss of flavonoids in fresh-cut onion slices due to washing with sodium hypochlorite (Pérez-Gregorio *et al.*, 2011); induced browning of lettuce due to gaseous ClO<sub>2</sub> (Mahmoud *et al.*, 2008); color alterations of red bell peppers and strawberries due to sanitization with H<sub>2</sub>O<sub>2</sub> solution (Alexandre *et al.*, 2012).

Several other studies involving ozonation have reported a significant loss in vitamin C content. Beltrán *et al.* (2005) reported a loss of ascorbic acid in lettuce when ozonized water at 20 mg of O<sub>3</sub>/L was used. Tiwari *et al.* (2009) reported the same reduction and also a loss of anthocyanin in strawberry juice bubbled with 7.8% of O<sub>3</sub> for 10 min. A loss of ascorbic acid as well as carotenoids was reported by Chauhan *et al.* (2011) when they used ozonized water to wash carrots for 10 min.

On the other hand, Karaca and Velioglu (2014) used 12 mg/L of ozonized water for 15 min to wash lettuce, spinach and parsley but did not observe any changes in the levels of chlorophyll, ascorbic acid, total phenolic contents or antioxidant activity. Aguayo *et al.* (2013) also did not reported changes in the quality of tomato slices when 0.4 mg/L of ozonized water was applied for 3 min. Tzortzakis *et al.* (2007) did not observed changes in the quality of tomatoes stored for 6 days at 1 µmol/mol of O<sub>3</sub>.

Positive changes may also occur, as reported by Ali *et al.* (2014) when stored papaya fruit was

ozonized with O<sub>3</sub> at 2.5 ppm for 10 days. Higher total solid values, ascorbic acid, β-carotene, lycopene and antioxidant activity were obtained in relation to the control sample, meaning a lower decay in these compounds with time. Similar results were reported by Yeoh *et al.* (2014), when fresh-cut papaya was treated with O<sub>3</sub> at 9.2 mg/L for 20 min. The authors observed that the total phenolic content increased by 10.3%, which occurred due to the activation of certain enzymes that are stimulated by different abiotic stresses.

When applied to cereal grains, the effects of O<sub>3</sub> may be confirmed through the changes of certain quality parameters in the flour obtained from the ozonized grains. Violleau *et al.* (2012) applied 5 g of O<sub>3</sub>/kg grains and reported greater force and less extensibility in dough due to the oxidation of gluten, which interfered in the technological properties of the flour. Li *et al.* (2012) used 5 g of O<sub>3</sub> for 60 min and reported greater dough development time and an increased stability in the flour, which also occurs when other oxidizing agent, such as potassium bromate or chlorine are used. In this case, O<sub>3</sub> has the advantage of promoting the same effects, but without leaving potassium bromate or chlorine residues in the food. Sandhu *et al.* (2011) reported similar changes when they exposed wheat flour (100 g) to 1500 mg of O<sub>3</sub> for 45 min.

Sandhu *et al.* (2012) described that the use of O<sub>3</sub> at 1500 mg/kg for 45 min on wheat flour resulted in depolymerization of high molecular weight amylopectin, with a consequent increase in low molecular weight polymers, which may be useful for flours with low viscosity, high clarity, and low temperature stability requirements.

As occurs in fruits rich in pigments, ozonation may also react with the conjugated double bonds in the carotenoid of wheat flour decreasing the yellowness (Sandhu *et al.*, 2011). Li *et al.* (2012a) reported this effect when they used 5 g of O<sub>3</sub>/h for 60 min, and suggested that ozone can be used as a bleaching agent for wheat flour.

An undesirable effect of O<sub>3</sub> reported in some studies, is the formation of an unpleasant smell in the flour after the ozonation process, due to the formation of volatile low molecular weight compounds, as described by Chittrakorn *et al.* (2014) and Li *et al.* (2012). However, according to these authors, the aeration of the flour using storage ventilation can easily eliminate this problem.

In animal products, O<sub>3</sub> can also be applied to control microorganisms without promoting unwanted changes in quality, as reported by Iacumin *et al.* (2012) when sausages were stored in at 1 ppm atmosphere of

O<sub>3</sub>. Kamotani *et al.* (2010) used gaseous O<sub>3</sub> (9.7% in oxygen for 40 min) for processing eggs as an alternative treatment to pasteurization and reported no significant changes in the characteristics of the product, and with the advantage of not having to use heat. On the other hand, Uzun *et al.* (2012) observed reduced solubility of whey protein isolates and egg yolk proteins as a consequence of both aqueous and gas ozonation. Ozone treatment also negatively affected the emulsion activity of whey protein isolates and reduced their stability.

#### *Perspectives for the use of O<sub>3</sub> in food processing*

Based on the recent evolution of studies involving ozonation of food, more research can be expected with the goal to integrate gas ozonation or ozonized water into the traditional or innovative food processing procedures in order to reduce residues and contaminants, especially pesticide residues and mycotoxins. In fact, some studies have already been done, demonstrating interesting results. Chauhan *et al.* (2011) studied the washing process of carrots using ozonized water followed by storage in a controlled atmosphere, which reduced the lignification and kept the quality of fresh-cut carrots for 30 days. Chen *et al.* (2013) evaluated the effects of ultrasound combined with ozonized water on the degradation of organophosphorus pesticides residues in lettuce. With the use of both technologies the average levels of reduction peaked 82%, without negatively affecting the quality of the vegetable. Puzyr *et al.* (2010) studied the efficacy of ozonation followed by adsorption on nanodiamonds hydrosol to decrease the aflatoxin B1 content. These authors suggested that the use of these technologies together is a new approach to mycotoxin decontamination, where the ozonation degrades the toxin and the nanodiamond adsorbs the residual levels with high efficacy. Dudziak (2012) evaluated the application of an integrated system of ozonation and nanofiltration using a cellulose acetate membrane to remove the mycotoxin zearalenone in water. This study showed that with the combined use of these technologies it was possible to eliminate 100% of the zearalenone.

Ozone has a significant potential to be applied as a substitute of the normal chemical agents used in vegetable sanitization, fumigation of grains and in food storage, especially as it does not leave residues due to the treatment. Special attention should be given to the processing of organic food. According to the United States Department of Agriculture, organic food can be treated with ozonation, and the food can be classified as “100% organic” or “organic”, depending on the O<sub>3</sub> usage (USDA, 2011). Other

regulatory agencies in different countries do not make restrictions on the use of O<sub>3</sub> as a sanitizing agent for organic food, which makes its use very promising in this sector.

Due to the consumer's interest in new food processing technologies and the excellent results promoted by ozonation in improving the quality of food, new types of ozonizers are expected to appear on the market not only for industry but also for domestic use. Based on such expectations, more scientific research should be conducted, evaluating the effects of ozonation on the removal of different residues and contaminants, the possible formation of toxic degradation products and, also, the processing cost studies in order to disseminate a more practical and commercial application of this technology.

## Conclusion

Current consumer perceptions concerning food safety and the rising concern about the presence of residues and contaminants has opened new fields of study for emerging food processing technologies. Gaseous ozonation and ozonized water are interesting nonthermal methods and with high efficacy for the decontamination of pesticide residues and mycotoxins in different types of foods. According to recent research, O<sub>3</sub> can degrade and reduce both mycotoxins and pesticide residues in food. As verified by mass spectrometry, using structure–activity relationships and also according to the studies with animal models, the toxic effects of food contaminated by mycotoxins can be eliminated or significantly reduced using ozonation processes. Some negative effects of ozonation are the undesirable changes that may occur in food quality, such as, a loss of phenolic compounds and ascorbic acid, inactivation of some enzymes and changes in color, especially when applied to fresh vegetables and fruit products. However, when optimal conditions are determined for each food, these effects will be greatly reduced. More studies are needed to clarify in depth the effects of ozonation on a higher number of mycotoxins and pesticides as well as the influence of the process on a greater variety of foods. The most recent studies that have demonstrated the excellent effects of O<sub>3</sub> in improving the quality of food should result in greater interest of this technology by the food industry, and consequently a wider acceptance and popularization of ozonized products by consumers.

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## References

- Aguayo, E., Escalona, V., Silveira, A. C. and Artés, F. 2013. Quality of tomato slices disinfected with ozonated water. *Food Science and Technology International* 20(3): 227-235.
- Akbas, M. Y. and Ozdemir, M. 2006. Effect of different ozone treatments on aflatoxin degradation and physicochemical properties of pistachios. *Journal of the Science of Food and Agriculture* 86(13): 2099–2104.
- Akoto, O., Oppong-Otoo, J. and Osei-Fosu, P. 2015. Carcinogenic and non-carcinogenic risk of organochlorine pesticide residues in processed cereal-based complementary foods for infants and young children in Ghana. *Chemosphere* 132: 193–199.
- Alencar, E. R., Faroni, L. R. D., Soares, N. F. F., Silva, W. A. and Carvalho, M. C. S. 2012. Efficacy of ozone as a fungicidal and detoxifying agent of aflatoxins in peanuts. *Journal of the Science of Food and Agriculture* 92(4): 899–905.
- Alexandre, E.M.C., Brandão, T.R.S., Silva, C.L.M. 2012. Assessment of the impact of hydrogen peroxide solutions on microbial loads and quality factors of red bell peppers, strawberries and watercress. *Food Control* 27(1): 362–368.
- Ali, A., Ong, M. K. and Forney, C. F. 2014. Effect of ozone pre-conditioning on quality and antioxidant capacity of papaya fruit during ambient storage. *Food Chemistry* 142: 19–26.
- Antonissen, G., Martel, A., Pasmans, F., Ducatelle, R., Verbrugge, E., Vandenbroucke, V. and Croubels, S. 2014. The impact of *fusarium* mycotoxins on human and animal host susceptibility to infectious diseases. *Toxins* 6(2): 430–452.
- Bargańska, Ż., Ślebioda, M. and Namieśnik, J. 2013. Pesticide residues levels in honey from apiaries located of Northern Poland. *Food Control* 31(1): 196–201.
- Beltrán, D., Selma, M. V, Marín, A. and Gil, M. I. 2005. Ozonated water extends the shelf life of fresh-cut lettuce. *Journal of Agricultural and Food Chemistry* 53(14): 5654–63.
- Blaznik, U., Yngve, A., Eržen, I. and Hlastan Ribič, C. 2015. Consumption of fruits and vegetables and probabilistic assessment of the cumulative acute exposure to organophosphorus and carbamate pesticides of schoolchildren in Slovenia. *Public Health Nutrition*, 20: 1–7.
- Caballero, P. a., Gómez, M. and Rosell, C. M. 2007. Bread quality and dough rheology of enzyme-supplemented wheat flour. *European Food Research and Technology* 224(5): 525–534.
- Cataldo, F. 2008. Ozone Decomposition of Patulin—A

- Micotoxin and food contaminant. *Ozone: Science & Engineering* 30(3): 197–201.
- Chauhan, O. P., Raju, P. S., Ravi, N., Singh, A. and Bawa, A. S. 2011. Effectiveness of ozone in combination with controlled atmosphere on quality characteristics including lignification of carrot sticks. *Journal of Food Engineering* 102(1): 43–48.
- Cheli, F., Battaglia, D., Gallo, R. and Dell'Orto, V. 2014. EU legislation on cereal safety: An update with a focus on mycotoxins. *Food Control* (37): 315–325.
- Chen, J. Y., Lin, Y. J. and Kuo, W. C. 2013. Pesticide residue removal from vegetables by ozonation. *Journal of Food Engineering* 114(3): 404–411.
- Chen, R., Ma, F., Li, P.W., Zhang, W., Ding, X.X., Zhang, Q. and Xu, B.C. 2014. Effect of ozone on aflatoxins detoxification and nutritional quality of peanuts. *Food Chemistry* 146: 284–288.
- Chittrakorn, S., Earls, D. and MacRitchie, F. 2014. Ozonation as an alternative to chlorination for soft wheat flours. *Journal of Cereal Science* 60(1): 217–221.
- Chiu, Y. H., Afeiche, M. C., Gaskins, A. J., Williams, P. L., Petrozza, J. C., Tanrikut, C. and Havarro, J. E. 2015. Fruit and vegetable intake and their pesticide residues in relation to semen quality among men from a fertility clinic. *Human Reproduction* 30(6): 1342–1351.
- Chourasiya, S., Khillare, P. S. and Jyethi, D. S. 2015. Health risk assessment of organochlorine pesticide exposure through dietary intake of vegetables grown in the periurban sites of Delhi, India. *Environmental Science and Pollution Research* 22(8): 5793–5806.
- De, A., Bose, R., Kumar, A. and Mozumdar, S. 2014. Targeted delivery of pesticides using biodegradable polymeric nanoparticles. *Fundamentals and Applications of Controlled Release Drug Delivery*. New Delhi: Springer India, 99p.
- De Ruyck, K., De Boevre, M., Huybrechts, I. and De Saeger, S. 2015. Dietary mycotoxins, co-exposure, and carcinogenesis in humans: Short review. *Mutation Research/Reviews in Mutation Research*. In press.
- Dewan, P., Jain, V., Gupta, P. and Banerjee, B. D. 2013. Organochlorine pesticide residues in maternal blood, cord blood, placenta, and breastmilk and their relation to birth size. *Chemosphere* 90(5): 1704–1710.
- Diao, E., Hou, H., Chen, B., Shan, C. and Dong, H. 2013. Ozonolysis efficiency and safety evaluation of aflatoxin B<sub>1</sub> in peanuts. *Food and Chemical Toxicology* 55: 519–25.
- Diao, E., Shan, C., Hou, H., Wang, S., Li, M. and Dong, H. 2012. Structures of the ozonolysis products and ozonolysis pathway of aflatoxin B<sub>1</sub> in acetonitrile solution. *Journal of Agricultural and Food Chemistry* 60(36): 9364–70.
- Dudziak, M. 2012. Removal of zearalenone from water by means of ozonation and integrated system of ozonation/nanofiltration. *Ecol chem eng* 19(07): 779–785.
- EFSA. European Food Safety Authority. 2013a. Aflatoxins (sum of B<sub>1</sub>, B<sub>2</sub>, G<sub>1</sub>, G<sub>2</sub>) in cereals and cereal-derived food products. European Food Safety Authority Technical Report. Parma, Italy. Downloaded from <http://www.efsa.europa.eu/en/supporting/doc/406e.pdf> on 9/2/2015.
- EFSA. European Food Safety Authority. 2013b. Deoxynivalenol in food and feed: occurrence and exposure. *EFSA Journal* 11(10): 3379–3434.
- EFSA. 2015. European Food Safety Authority. The 2013 European Union Report on Pesticide Residues in Food. *EFSA Journal* 13(3): 169.
- El-Desouky, T., Sharoba, A., Desouky, A. I. E. and El-Mansy, H. 2012. Effect of Ozone Gas on Degradation of Aflatoxin B<sub>1</sub> and *Aspergillus flavus* gungal. *Journal of Environmental & Analytical Toxicology* 02(02): 1–6.
- El-Morsi, D. and Rahman, R. H. A. 2012. Pesticides Residues in Egyptian Diabetic Children: A Preliminary Study. *Journal of Clinical Toxicology* 02(06): 1–6.
- FAO. Food and Agriculture Organization. 2001. Joint FAO/WHO Food Standards Programme. Definitions for the Purposes of the Codex Alimentarius, 5–8.
- FDA. Food and Drug Administration. 2001. Secondary Direct Food Additives Permitted in Food for Human Consumption. *Federal Register* 66(123): 33829–33830.
- FDA. Food and Drug Administration. 2014. U.S. Food and Drug Administration. Sec. Title 21. 801.415. Maximum acceptable level of ozone. *Code of Federal Regulations*, 8.
- Feliziani, E., Romanazzi, G. and Smilanick, J. L. 2014. Application of low concentrations of ozone during the cold storage of table grapes. *Postharvest Biology and Technology* 93: 38–48.
- Freitas-Silva, O. and Venâncio, A. 2010. Ozone applications to prevent and degrade mycotoxins: a review. *Drug Metabolism Reviews* 42(4): 612–620.
- Freitas-Silva, O. 2011. Mycotoxins and mycobiota in Brazil nuts and strategies for their control. (Dissertation) Universidade do Minho.
- Gabler, F. M., Smilanick, J. L., Mansour, M. F. and Karaca, H. 2010. Influence of fumigation with high concentrations of ozone gas on postharvest gray mold and fungicide residues on table grapes. *Postharvest Biology and Technology* 55(2): 85–90.
- Gaou, I., Dubois, M., Pfohl-Leszkowicz, A., Coste, C., De Jouffrey, S. and Parent-Massin, D. 2005. Safety of Oxygreen®, an ozone treatment on wheat grains. Part 1. A four-week toxicity study in rats by dietary administration of treated wheat. *Food Additives and Contaminants* 22(11): 1113–1119.
- Gerding, J., Ali, N., Schwartzbord, J., Cramer, B., Brown, D. L., Degen, G. H. and Humpf, H. U. 2015. A comparative study of the human urinary mycotoxin excretion patterns in Bangladesh, Germany, and Haiti using a rapid and sensitive LC-MS/MS approach. *Mycotoxin Research* 31(3):127-136.
- Giordano, B. N., Simao, V., Manfio, D., Galvao, S., Scussel, J. N. and Scussel, V. 2010. Reduction of in-shell Brazil nut (*Bertholletia excelsa* H.B.K.) aflatoxin contamination by ozone gas application during storage. In 10th International Working Conference

- on Stored Product Protection Reduction. Estoril. Downloaded from [pub.jki.bund.de/index.php/JKA/article/download/550/1265](http://pub.jki.bund.de/index.php/JKA/article/download/550/1265) on 9/2/2015
- Giordano, B. N. E., Nones, J. and Scussel, V. M. 2012. Susceptibility of the in-shell Brazil nut mycoflora and aflatoxin contamination to ozone gas treatment during Storage. *Journal of Agricultural Science* 4(8): 1–10.
- Glaser, N. and Stopper, H. 2012. Patulin: Mechanism of genotoxicity. *Food and Chemical Toxicology* 50(5): 1796–1801.
- Gorayeb, T. C. C., Casciatori, F. P., Bianchi, V. L. Del, and Thoméo, J. C. 2009. HACCP plan proposal for a typical Brazilian peanut processing company. *Food Control* 20(7): 671–676.
- Greene, A. K., Guzel-seydim, Z. B. and Seydim, A. C. 2012. Chemical and physical properties of ozone. In C. O'Donnell, B. K. Tiwari, P. J. Cullen, and R. G. Rice (Eds.), *Ozone in Food Processing* (pp. 19–32). John Wiley and Sons.
- Handford, C. E., Elliott, C. T. and Campbell, K. 2015. A review of the global pesticide legislation and the scale of challenge in reaching the global harmonization of food safety standards. *Integrated Environmental Assessment and Management*, 9999(9999): 1-12.
- Hansen, L. S., Hansen, P. and Vagn Jensen, K.-M. 2013. Effect of gaseous ozone for control of stored product pests at low and high temperature. *Journal of Stored Products Research* 54: 59–63.
- Iacumin, L., Manzano, M. and Comi, G. 2012. Prevention of *Aspergillus ochraceus* growth on and Ochratoxin a contamination of sausages using ozonated air. *Food Microbiology* 29(2): 229–32.
- IARC. International Agency for Research on Cancer. 2002. Monographs on the Evaluation of Carcinogenic Risks to Humans. Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. Downloaded from <http://monographs.iarc.fr/ENG/Monographs/vol82/> on 9/2/2015.
- Ibanoğlu, Ş. 2001. Influence of tempering with ozonated water on the selected properties of wheat flour. *Journal of Food Engineering* 48(4): 345–350.
- Iha, M. H., Barbosa, C. B., Heck, A. R. and Trucksess, M. W. 2014. Aflatoxin M1 and ochratoxin A in human milk in Ribeirão Preto-SP, Brazil. *Food Control* 40: 310–313.
- Ikehata, K., Jodeiri Naghashkar, N. and Gamal El-Din, M. 2006. Degradation of aqueous pharmaceuticals by ozonation and advanced oxidation processes: A Review. *Ozone: Science & Engineering* 28(6): 353–414.
- Ikeura, H., Hamasaki, S. and Tamaki, M. 2013. Effects of ozone microbubble treatment on removal of residual pesticides and quality of persimmon leaves. *Food Chemistry* 138(1): 366–371.
- Ikeura, H., Kobayashi, F. and Tamaki, M. 2011. Removal of residual pesticide, fenitrothion, in vegetables by using ozone microbubbles generated by different methods. *Journal of Food Engineering* 103(3): 345–349.
- Ikeura, H., Kobayashi, F. and Tamaki, M. 2011. Removal of residual pesticides in vegetables using ozone microbubbles. *Journal of Hazardous Materials* 186(1): 956–9.
- Inan, F., Pala, M. and Doymaz, I. 2007. Use of ozone in detoxification of aflatoxin B<sub>1</sub> in red pepper. *Journal of Stored Products Research* 43(4): 425–429.
- Jardim, A. N. O., Mello, D. C., Goes, F. C. S., Frota, E. F. and Caldas, E. D. 2014. Pesticide residues in cashew apple, guava, kaki and peach: GC-μECD, GC-FPD and LC-MS/MS multiresidue method validation, analysis and cumulative acute risk assessment. *Food Chemistry* 164: 195–204.
- Kamotani, S., Hooker, N., Smith, S. and Lee, K. 2010. Consumer acceptance of ozone-treated whole shell eggs. *Journal of Food Science* 75(2): 103–107.
- Karaca, H. and Velioglu, Y. S. 2014. Postharvest biology and technology effects of ozone treatments on microbial quality and some chemical properties of lettuce, spinach, and parsley. *Postharvest Biology and Technology* 88: 46–53.
- Kaushik, G., Satya, S. and Naik, S. N. 2009. Food processing a tool to pesticide residue dissipation – A review. *Food Research International*, 42(1): 26–40.
- Kher, S. V., De Jonge, J., Wentholt, M. T. A., Deliza, R., de Andrade, J. C., Cnossen, H. J., Frewer, L. J. 2013. Consumer perceptions of risks of chemical and microbiological contaminants associated with food chains: a cross-national study. *International Journal of Consumer Studies* 37(1): 73–83.
- Kusvuran, E., Yildirim, D., Mavruk, F. and Ceyhan, M. 2012. Removal of chlorpyrifos ethyl, tetradifon and chlorothalonil pesticide residues from citrus by using ozone. *Journal of Hazardous Materials* 241-242, 287–300.
- Lafi, W. K. and Al-Qodah, Z. 2006. Combined advanced oxidation and biological treatment processes for the removal of pesticides from aqueous solutions. *Journal of Hazardous Materials* 137(1): 489–97.
- Li, M. M., Guan, E. Q. and Bian, K. 2014. Effect of ozone treatment on deoxynivalenol and quality evaluation of ozonised wheat. *Food Additives & Contaminants: Part A* 32(4): 544–553.
- Li, M., Zhu, K., Wang, B., Guo, X., Peng, W. and Zhou, H. 2012a. Evaluation the quality characteristics of wheat flour and shelf-life of fresh noodles as affected by ozone treatment. *Food Chemistry* 135(4): 2163–2169.
- Li, Y., Zhou, Y.-C., Yang, M.-H. and Ou-Yang, Z. 2012b. Natural occurrence of citrinin in widely consumed traditional Chinese food red yeast rice, medicinal plants and their related products. *Food Chemistry* 132(2): 1040–1045.
- Liu, Y., Pan, X. and Li, J. 2015. Current agricultural practices threaten future global food production. *Journal of Agricultural and Environmental Ethics* 28(2): 203–216.
- Lombard, M. J. 2014. Mycotoxin exposure and infant and young child growth in Africa: What do we know? *Annals of Nutrition and Metabolism* 64: 42–52.
- Lozowicka, B. 2015. Health risk for children and adults consuming apples with pesticide residue. *Science of The Total Environment* 502: 184–198.

- Lozowicka, B., Kaczynski, P., Paritova, A. E., Kuzembekova, G. B., Abzhalieva, A. B., Sarsembayeva, N. B. and Alihan, K. 2014. Pesticide residues in grain from Kazakhstan and potential health risks associated with exposure to detected pesticides. *Food and Chemical Toxicology* 64: 238–248.
- Luo, X., Wang, R., Wang, L., Li, Y., Zheng, R., Sun, X. and Tao, G. 2014a. Analyses by UPLC Q-TOF MS of products of aflatoxin B<sub>1</sub> after ozone treatment. *Food Additives & Contaminants. Part A* 31(1): 105–10.
- Luo, X., Wang, R., Wang, L., Li, Y., Wang, Y. and Chen, Z. 2014b. Detoxification of aflatoxin in corn flour by ozone. *Journal of the Science of Food and Agriculture* 94(11): 2253-2258.
- Luo, X., Wang, R., Wang, L., Li, Y., Bian, Y. and Chen, Z. 2014c. Effect of ozone treatment on aflatoxin B1 and safety evaluation of ozonized corn. *Food Control* 37(1): 171–176.
- Magnussen, A. 2013. Aflatoxins, hepatocellular carcinoma and public health. *World Journal of Gastroenterology* 19(10): 1508.
- Malik, A. K., Blasco, C. and Picó, Y. 2010. Liquid chromatography–mass spectrometry in food safety. *Journal of Chromatography A* 1217(25): 4018–4040.
- Mahmoud, B.S.M., Vaidya, N.A., Corvalan, C.M. and Linton, R.H. 2008. Inactivation kinetics of inoculated *Escherichia coli* O157:H7, *Listeria monocytogenes* and *Salmonella* Poona on whole cantaloupe by chlorine dioxide gas. *Food Microbiology* 25 (1): 857–865.
- Marroquín-Cardona, A. G., Johnson, N. M., Phillips, T. D. and Hayes, A. W. 2014. Mycotoxins in a changing global environment - A review. *Food and Chemical Toxicology* 69: 220-230.
- McClurkin, J. D., Maier, D. E. and Ileleji, K. E. 2013. Half-life time of ozone as a function of air movement and conditions in a sealed container. *Journal of Stored Products Research* 55: 41–47.
- McDonough, M. X., Campabadal, C. A., Mason, L. J., Maier, D. E., Denvir, A. and Woloshuk, C. 2011. Ozone application in a modified screw conveyor to treat grain for insect pests, fungal contaminants, and mycotoxins. *Journal of Stored Products Research* 47(3): 249–254.
- Min, Z. W., Hong, S.M., Yang, I.C., Kwon, H.Y., Kim, T.K. and Kim, D.H. 2012. Analysis of pesticide residues in brown rice using modified QuEChERS multiresidue method combined with electrospray ionization-liquid chromatography-tandem mass spectrometric detection. *Journal of the Korean Society for Applied Biological Chemistry* 55(6): 769–775.
- Mostafalou, S. and Abdollahi, M. 2013. Pesticides and human chronic diseases: Evidences, mechanisms, and perspectives. *Toxicology and Applied Pharmacology* 268(2): 157–177.
- O'Donnell, C., Tiwari, B. K., Cullen, P. J. and Rice, R. G. 2012. Status and trends of ozone in food processing. In C. O'Donnell, B. K. Tiwari, P. J. Cullen and R. G. Rice (Eds.), *Ozone in Food Processing*. Oxford, UK: Wiley-Blackwell. 312 p.
- Oliveira, L. A. B., Pacheco, H. P. and Scherer, R. 2016. Flutriafol and pyraclostrobin residues in Brazilian green coffees. *Food Chemistry* 190: 60–63.
- Ormad, M. P., Miguel, N., Claver, A., Matesanz, J. M. and Ovelheiro, J. L. 2008. Pesticides removal in the process of drinking water production. *Chemosphere* 71(1): 97–106.
- Parrón, T., Requena, M., Hernández, A. F. and Alarcón, R. 2014. Environmental exposure to pesticides and cancer risk in multiple human organ systems. *Toxicology Letters* 230(2): 157–165.
- Patil, S., Torres, B., Tiwari, B. K., Wijngaard, H. H., Bourke, P., Cullen, P. J. and Valdramidis, V. P. 2010. Safety and quality assessment during the ozonation of cloudy apple juice. *Journal of Food Science* 75(7): 437–43.
- Pérez-Gregorio, M.R., González-Barreiro, C., Rial-Otero, R. and Simal-Gándara, J. 2011. Comparison of sanitizing technologies on the quality appearance and antioxidant levels in onion slices. *Food Control* 22(1): 2052–2058.
- PubChem. 2015. National Center for Biotechnology Information. PubChem Compound Database; CID=24823. Downloaded from <https://pubchem.ncbi.nlm.nih.gov/compound/24823> on 9/2/2015.
- Puzyr', A. P., Burov, A. E., Bondar', V. S. and Trusov, Y. N. 2010. Neutralization of aflatoxin B<sub>1</sub> by ozone treatment and adsorption by nanodiamonds. *Nanotechnologies in Russia* 5(1-2): 137–141.
- Rocha, E. B., Freire, F.C. O., Maia, F. E. F., Guedes, M. I. F., Rondina, D., Rocha, E. B. M. and Guedes, I. F. 2014. Mycotoxins and their effects on human and animal health. *Food Control* 36(1): 159–165.
- Rodríguez López, D., Ahumada, D. A., Díaz, A. C. and Guerrero, J. A. 2014. Evaluation of pesticide residues in honey from different geographic regions of Colombia. *Food Control* 37(1): 33–40.
- Romero-González, R. 2015. Food safety: how analytical chemists ensure it. *Anal. Methods*, 7(17): 7193–7201.
- Rubert, J., León, N., Sáez, C., Martins, C. P. B., Godula, M., Yusà, V. and Soler, C. 2014. Evaluation of mycotoxins and their metabolites in human breast milk using liquid chromatography coupled to high resolution mass spectrometry. *Analytica Chimica Acta*, 820: 39–46.
- Sandhu, H. P. S., Manthey, F. A. and Simsek, S. 2011. Quality of bread made from ozonated wheat (*Triticum aestivum* L.) flour. *Journal of the Science of Food and Agriculture* 91(9): 1576–1584.
- Sandhu, H. P. S. S., Manthey, F. A. and Simsek, S. 2012. Ozone gas affects physical and chemical properties of wheat (*Triticum aestivum* L.) starch. *Carbohydrate Polymers* 87(2): 1261–1268.
- Savi, G. D., Piacentini, K. C., Bittencourt, K. O. and Scussel, V. M. 2014. Ozone treatment efficiency on *Fusarium graminearum* and deoxynivalenol degradation and its effects on whole wheat grains (*Triticum aestivum* L.) quality and germination. *Journal of Stored Products Research* 59: 245–253.
- Savi, G. D., Piacentini, K. C. and Scussel, V. M. 2014.

- Ozone treatment efficiency in *Aspergillus* and *Penicillium* growth inhibition and mycotoxin degradation of stored wheat grains (*Triticum aestivum* L.). *Journal of Food Processing and Preservation*.
- Savi, G. D., Piacentini, K. C. and Scussel, V. M. 2015. Reduction in residues of deltamethrin and fenitrothion on stored wheat grains by ozone gas. *Journal of Stored Products Research*, 61: 65–69.
- Sorrenti, V., Di Giacomo, C., Acquaviva, R., Barbagallo, I., Bognanno, M. and Galvano, F. 2013. Toxicity of Ochratoxin A and Its Modulation by Antioxidants: A Review. *Toxins* 5(10): 1742–1766.
- Srey, C., Kimanya, M. E., Routledge, M. N., Shirima, C. P. and Gong, Y. Y. 2014. Deoxynivalenol exposure assessment in young children in Tanzania. *Molecular Nutrition & Food Research* 58(7): 1574–1580.
- Stoev, S. D. 2015. Foodborne mycotoxicoses, risk assessment and underestimated hazard of masked mycotoxins and joint mycotoxin effects or interaction. *Environmental Toxicology and Pharmacology* 39(2): 794–809.
- Takahashi, M., Chiba, K. and Li, P. 2007. Formation of hydroxyl radicals by collapsing ozone microbubbles under strongly acidic conditions. *The Journal of Physical Chemistry B* 111(39): 11443–11446.
- Tapp, C. and Rice, R. G. 2012. Generation and control of ozone. In C. O'Donnell, B. K. Tiwari, P. J. Cullen, and R. G. Rice (Eds.), *Ozone in Food Processing*. Oxford, UK: Wiley-Blackwell. 312 p.
- Tibola, C. S., Fernandes, J. M. C., Guarienti, E. M. and Nicolau, M. 2015. Distribution of *Fusarium* mycotoxins in wheat milling process. *Food Control* 53: 91–95.
- Tiwari, B. K., O'Donnell, C. P., Patras, A., Brunton, N. and Cullen, P. J. 2009. Effect of ozone processing on anthocyanins and ascorbic acid degradation of strawberry juice. *Food Chemistry* 113(4): 1119–1126.
- Tzortzakis, N., Borland, A., Singleton, I. and Barnes, J. 2007. Impact of atmospheric ozone-enrichment on quality-related attributes of tomato fruit. *Postharvest Biology and Technology* 45(3): 317–325.
- U.S.EPA - Environmental Protection Agency. 2014. Pesticides: Health and Safety. Downloaded from <http://www.epa.gov/pesticides/health/human.htm> on 9/2/2015.
- USDA. U.S. Department of Agriculture. 2011. National Organic Program (NOP). Agricultural Marketing Service. Nonagricultural (nonorganic) substances allowed as ingredients in or on processed products labeled as “organic” or “made with organic”. Downloaded from <http://www.gpo.gov/fdsys/pkg/CFR-2011-title7-vol3/pdf/CFR-2011-title7-vol3-sec205-605.pdf> on 9/2/2015.
- Uzun, H., Ibanoglu, E., Catal, H. and Ibanoglu, S. 2012. Effects of ozone on functional properties of proteins. *Food Chemistry* 134(2): 647–654.
- Violleau, F., Pernot, A. G. and Surel, O. 2012. Effect of Oxygreen wheat ozonation process on bread dough quality and protein solubility. *Journal of Cereal Science* 55(3): 392–396.
- WCBBC. Workers' Compensation Board of British Columbia. 2006. Library and Archives Canada Cataloguing in Publication Data. Ozone safe work practices: WCBBC.
- WHO. World Health Organization. 2006. Air quality guidelines for particulate matter, ozone, nitrogen dioxide and sulfur dioxide: global update 2005. Geneva: World Health Organization, 1–22. Downloaded from [http://whqlibdoc.who.int/hq/2006/WHO\\_SDE\\_PHE\\_OEH\\_06.02\\_eng.pdf?ua=1](http://whqlibdoc.who.int/hq/2006/WHO_SDE_PHE_OEH_06.02_eng.pdf?ua=1) on 9/2/2015
- Wu, F., Groopman, J. D. and Pestka, J. J. 2014. Public health impacts of foodborne mycotoxins. *Annual Review of Food Science and Technology* 5(1): 351–372.
- Wu, J. G., Luan, T. G., Lan, C. Y., Lo, W. H. and Chan, G. Y. S. 2007. Efficacy evaluation of low-concentration of ozonated water in removal of residual diazinon, parathion, methyl-parathion and cypermethrin on vegetable. *Journal of Food Engineering* 79(3): 803–809.
- Yazar, S. and Omurtag, G. Z. 2008. Fumonisin, Trichothecenes and Zearalenone in Cereals. *International Journal of Molecular Sciences* 9(11): 2062–2090.
- Yeoh, W. K., Ali, A., Forney, C. F. and Keat, W. 2014. Effects of ozone on major antioxidants and microbial populations of fresh-cut papaya. *Postharvest Biology and Technology* 89: 56–58.
- Young, J. C., Zhu, H. and Zhou, T. 2006. Degradation of trichothecene mycotoxins by aqueous ozone. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association* 44(3): 417–24.
- Yusa, V., Millet, M., Coscolla, C. and Roca, M. 2015. Analytical methods for human biomonitoring of pesticides. A review. *Analytica Chimica Acta*. In Press.
- Zorlugenç, B., Kiroğlu Zorlugenç, F., Öztekin, S. and Evliya, I. B. 2008. The influence of gaseous ozone and ozonated water on microbial flora and degradation of aflatoxin B1 in dried figs. *Food and Chemical Toxicology* 46(12): 3593–3597.